

Evaluation of some plant species for their resistance against root-knot nematode *Meloidogyne* spp.

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Abstract

The resistance of 4 cotton, 2 sesame, and 6 flax cultivars to the root-knot nematodes *Meloidogyne arenaria* (Ma), *M. incognita* race 1 (Mil) and/or *M. javanica* (Mj) was determined in green house studies. The results showed that the cotton cvs Giza 77, Giza 85, Giza 88, Giza 89 and sesame cvs Shandwil 1 and Toshka 1 were resistant (R) to the tested nematode species. On the other hand, six flax cultivars were tested, among them Giza 1 and Giza 5 were susceptible (S) to all the tested nematode species; Giza 6 was susceptible to Ma and Mj, while Sakha 1 was susceptible to Ma only. Giza 6 was highly susceptible (HS) to Mj only whereas Sakha 1 was highly susceptible to Mi and Mj. The flax cultivars Giza 7 and Giza 8 were susceptible to Ma and Mi while these were moderately susceptible (MS) to Mj only.

In three separate experiments the response of 4 banana cultivars (Baladi, Grand Naine, Magraby and Williams), 3 peach rootstocks (Bitter almond, Baladi, and Nemaguard), and 6 ornamental palm trees (California Washington palm, Mexican Washington, Canary date palm, Queen palm, Common palmetto, and Date palm) were tested for resistance to Mi1 and Mj. The banana and peach rootstock Bitter almond were highly susceptible to Mil and Mj. Baladi peach rootstock was highly susceptible to Mj. but resistant to Mil, whereas Nemaguard rootstock was resistant to both Mil and Mj. The tested palm trees viz., date palm, California and Mexican Washington palms were susceptible to Mil and Mj. whereas Canary date palm was moderately susceptible to both nematode species. The Queen palm was susceptible to Mil but moderately resistant to Mj. Common palmetto palm was moderately resistant to both Mil and Mj.

Keywords: Banana, cotton, flax, *Meloidogyne* spp., palm trees, pathogenicity peach rootstocks, resistance, sesame

In Egypt, plant parasitic nematodes are considered among the most important agricultural pests and the root-knot nematode *Meloidogyne* spp. is among the most important pests of many economic plant crops (Ibrahim, 1985; Ibrahim *et al.*, 2010; Ibrahim & Handoo, 2018). Previous studies showed that both *M. incognita* and *M. javanica* are of widespread occurrence and adversely affect the growth and production of many crops in northern Egypt; whereas *M.*

arenaria is less common and of limited occurrence (Ibrahim, 1985; Ibrahim *et al.*, 2010).

Screening plant cultivars for resistance to *Meloidogyne* spp., is of significance and can be useful in planning control measures for root-knot nematodes. Previous studies in Egypt showed differences in the tested plant cultivars in resistance to root-knot nematodes (Ibrahim & El-Saedy, 1982; Ibrahim & Handoo, 2018; Ibrahim *et al.*, 1982, 1983, 1999, 2013).

The objectives of this research were to study the resistance and susceptibility of certain cotton, flax, sesame and banana cultivars, peach rootstocks and palm trees to *M. arenaria*, *M. incognita* race 1 and/or *M. javanica*.

Materials and Methods

Inocula of the root-knot nematodes *M. arenaria* (Ma), *M. incognita* race 1 (Mil) and *M. javanica* (Mj) used in this study were obtained from infected roots of tomato (*Lycopersicon esculentum* Mill.) cultivar Rutgers grown in the greenhouse in the Faculty of Agriculture, Alexandria University, Alexandria, Egypt. Nematode eggs used in the pathogenicity tests were obtained from galled tomato roots using the method of Hussey & Barker (1973).

The reactions of four cotton (*Gossypium barbadense* L.) cultivars viz., Giza 77, Giza 85, Giza 88 and Giza 89; 6 flax (*Linum usitatissimum* L.) cultivars viz., Giza 1, Giza 5, Giza 6, Giza 7, Giza 8 and Sakha 1; sesame (*Sesamum indicum* L.) cultivars viz., Shandwil 1 and Toshka 1 to Ma, Mil and Mj were determined in three greenhouse experiments. Seeds of the tested plant cultivars were sown in 15-cm diameter plastic pots filled with 1 kg of equal portions of sterilized sand and clay soil. After emergence, seedlings were thinned to 2 seedlings/pot. Four weeks after emergence, soils of treated pots were inoculated with nematode eggs by creating holes near the plant roots and then adding initial population of 5000 eggs/pot of Ma, Mil or Mj. Untreated pots served as control. All the treatments were replicated five times and the experiments were repeated one time. Pots were arranged in a randomized complete block design in a greenhouse at 20-28° C.

In a separate greenhouse experiment, the reactions of four banana (*Musa sapientum* L.)

cultivars, the peach rootstocks, Bitter almond (*Prunus amygdalus* Batsch), peach (*P. persica* L.) cvs Balady and Nemaguard, and palm trees Queen palm (*Arecastrum romanzoffianum* (Cham.) Becc.), Canary date palm (*Phoenix canariensis* Chabaud), date palm (*Phoenix dactylifera* L.), Common palmetto (*Sabal palmetto* (Walt.) Lodd.), California Washington palm (*Washingtonia filifera* (Linden) Wendl) and Mexican Washington palm (*W. robustus* Wendl.) were tested for resistance to Mil and Mj. Two-month old seedlings of the tested banana cultivars, peach rootstocks and palm trees were transplanted in 20-cm diameter plastic pots filled with sterilized sand clay soil as one seedling/pot. Ten days after planting, soils of treated pots were inoculated with 5000 nematode eggs/pot of Mil or Mj, whereas uninoculated pots served as control. Treatments and controls were replicated five times and the experiments were repeated twice. Pots were arranged in randomized complete block design in a greenhouse at 20-28° C.

Experiments were terminated two months after soil inoculation and roots were washed free of soil. Roots were stained in aqueous solution of phloxine B (0.15g/1L) water for 15 minutes to detect the nematode egg-masses. The numbers of root-knot nematode galls and egg-masses were counted. Plants were rated according to the numbers of egg-masses observed on their roots. Plants with 0-2 egg-masses/plant were considered resistant, 3-10 egg-masses/plant moderately resistant, 11-30 egg-masses/plant moderately susceptible, 31-100 egg-masses/plant susceptible and < 100 egg-masses/plant highly susceptible (Taylor & Sasser, 1978).

Results and Discussion

Table 1 shows the reactions of the tested cotton, flax and sesame cultivars to infection with the root-knot nematodes; Ma, Mil and Mj.

Cotton cvs Giza 77, Giza 85, Giza 88, Giza 89 and sesame cvs Shandwill 1 and Toshka 1 have demonstrated resistance to the tested nematode populations. The present results agree with those of other authors who studied the effects of root-knot nematodes on cotton growth and yield and the resistance of cotton cultivars to *Meloidogyne* spp. (Ibrahim *et al.*, 1982; Starr & Martyn, 1991; Wrather *et al.*, 1992; Wendt & Greeff, 1991). On the other hand, flax cvs Giza 1, Giza 5, Giza 6, Giza 7, Giza 8 and Sakha 1 were either susceptible or highly susceptible to the tested nematode species, except cvs Giza 7 and Giza 8 were moderately susceptible to Mj (Table 1). These results confirm the findings of other workers (Abd El-Rahman *et al.*, 1986; Ivanova, 1984; Prasad & Khan, 1990; Zid, 1996).

The reactions of 4 banana cultivars, 3 peach rootstocks and 6 palm trees to Mil and Mj are presented in Table 2. Banana cvs Baladi, Grand Naine, Maghraby and Williams showed a highly susceptible reaction to both Mil and Mj. These results are similar to those of other studies that indicated the susceptibility of most banana cultivars to root-knot nematodes (Jabeen *et al.*, 1996; Jonathan *et al.*, 1999; Marcelion & Davide, 1967; Pinochet *et al.*, 1998; Sarah, 1989).

Data on the reactions of the tested peach rootstocks to Mil and Mj showed that Bitter almond was highly susceptible to both nematode species, and Baladi peach rootstock was highly susceptible to Mj but resistant to Mil whereas Nemaguard rootstock was resistant to both Mil and Mj (Table 2). These results are similar with those of previous studies on resistance of peach rootstocks to *Meloidogyne* species (Huettel & Hammerschlag, 1993; Marull & Pinochet, 1991; Marull *et al.*, 1991; Philis, 1989; Pinochet *et al.*, 1996).

The present results showed that date palm, California Washington palm and Mexican Washington palm were susceptible to Mil and Mj, where common palmetto palm was moderately resistant to Mil and Mj. The queen palm was susceptible to Mil but moderately resistant to Mj. Canary date palm was moderately susceptible to Mil and Mj (Table 2). The results confirm the findings of other workers who described the importance of plant parasitic nematodes on palm trees (Ibrahim *et al.*, 2000; Ismail & Eissa, 1993). Moreover, the status of date palm as a good host for *Meloidogyne* spp. reported by other authors (Ismail & Eissa, 1993; Youssef & Eissa, 1994) is supported by this study.

It is concluded from the study of determining the host reaction of certain cotton, flax, sesame and banana cultivars, peach rootstock and palm trees to *Meloidogyne arenaria*, *M. incognita* race1 and /or *M. javanica* that resistance to *Meloidogyne* spp. is of significance and can be useful to incorporate in breeding programs during planning control measures for root-knot nematodes.

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Table 1. Reaction of cotton, flax and sesame cultivars to infection with *Meloidogyne arenaria*, *M. incognita* and *M. javanica*.

Plant cultivars	Nematode species	No. of galls/plant	No. of egg-masses/plant	Reaction		
Cotton	<i>M. arenaria</i>	0*	0**	R		
	Giza 77	<i>M. incognita</i>	0	0	R	
		<i>M. javanica</i>	0	0	R	
		<i>M. arenaria</i>	0	0	R	
	Giza 85	<i>M. incognita</i>	0	0	R	
		<i>M. javanica</i>	0	0	R	
		<i>M. arenaria</i>	0	0	R	
	Giza 88	<i>M. incognita</i>	0	0	R	
		<i>M. javanica</i>	0	0	R	
		<i>M. arenaria</i>	0	0	R	
	Giza 89	<i>M. incognita</i>	0	0	R	
		<i>M. javanica</i>	0	0	R	
<i>M. arenaria</i>		0	0	R		
Flax	<i>M. arenaria</i>	102	85	S		
	Giza 1	<i>M. incognita</i>	114	95	S	
		<i>M. javanica</i>	110	85	S	
		<i>M. arenaria</i>	81	72	S	
	Giza 5	<i>M. incognita</i>	117	78	S	
		<i>M. javanica</i>	92	80	S	
		<i>M. arenaria</i>	105	74	S	
	Giza 6	<i>M. incognita</i>	146	80	S	
		<i>M. javanica</i>	176	139	HS	
		<i>M. arenaria</i>	85	68	S	
	Giza 7	<i>M. incognita</i>	109	56	S	
		<i>M. javanica</i>	52	13	MS	
		<i>M. arenaria</i>	76	61	S	
	Giza 8	<i>M. incognita</i>	121	78	S	
		<i>M. javanica</i>	52	13	MS	
		<i>M. arenaria</i>	102	80	S	
	Sakha 1	<i>M. incognita</i>	119	101	HS	
		<i>M. javanica</i>	136	121	HS	
		<i>M. arenaria</i>	0	0	R	
	Sesame	Shandwil 1	<i>M. incognita</i>	0	0	R
			<i>M. javanica</i>	0	0	R
<i>M. arenaria</i>			0	0	R	
Toshka 1		<i>M. incognita</i>	0	0	R	
		<i>M. javanica</i>	0	0	R	
		<i>M. arenaria</i>	0	0	R	

* Data is from one experiment each with five replications.

S = Susceptible, HS = Highly susceptible, MS = Moderately susceptible R= Resistant

Table 2. Reactions of four banana cultivars, three peach root-stocks and six palm trees to infection with *Meloidogyne incognita* and *M. javanica*

Test Plants	<i>M. incognita</i>			<i>M. javanica</i>		
	No. of galls/plant	No. of egg-masses/plant	Reaction	No. of galls/plant	No. of egg-masses/plant	Reaction
Banana						
Baladi	281*	143*	HS	407*	207*	HS
Grand Naine	918	614	HS	926	792	HS
Maghraby	805	543	HS	580	191	HS
Williams	308	116	HS	821	530	HS
Peach rootstocks						
Bitter almond	342	120	HS	364	134	HS
Balady peach	12	0	R	414	142	HS
Nemaguard peach	64	0	R	71	0	R
Palm trees						
Queen palm	53	48	S	11	9	MR
Canary date palm	39	29	MS	35	28	MS
Date palm	96	84	S	68	51	S
Common palmetto	5	3	MR	3	2	MR
California						
Washington palm	50	45	S	46	40	S
Mexican Washington palm	89	74	S	50	41	S

* Data is from one experiment each with five replications.

S = Susceptible, MS = Moderately susceptible, MR = Moderately resistant, R = Resistant

References

- Abd El-Rahman, H. F., Massoud, S. L. & Shohla, G. J. (1986). Screening and host suitability studies of five flax cultivars to the root-knot nematodes *Meloidogyne incognita*. *Faculty of Agriculture, Cairo University Bulletin*, 37, 541-549.
- Huettel, R. N. & Hammerschlag, F. A. (1993). Response of peach scion cultivars and rootstocks to *Meloidogyne incognita* in vitro and microplots. *Journal of Nematology*, 25, 472-475.
- Hussey, R. S. & Barker, K. R. (1973). A comparison methods of collecting inocula of *Meloidogyne* spp. including a new technique. *Plant Disease Reporter*, 54, 1025-1028.
- Ibrahim, I. K. A. (1985). The status of root-knot nematodes in the Middle East, Region VII of the International *Meloidogyne* Project. In: *Advanced Treatise on Meloidogyne. Vol. I, Biology and Control* (Ed. by) J. N. Sasser and C. C. Carter North Carolina State University Graphics. Raleigh, N.C., U.S.A.
- Ibrahim, I. K. A. & El-Saedy, M. A. (1982). Resistance of soybean cultivars to root-knot nematodes in Egypt. *Journal of Nematology*, 14, 447 (Abstr.).
- Ibrahim, I. K. A., Rezk, M. A. & Khalil, H. A. A. (1982). Reaction of fifteen malvaceous plant cultivars to root-knot nematodes, *Meloidogyne* spp. *Nematologia Mediterranea*, 10, 135-139.
- Ibrahim, I. K. A., Rezk, M. A. & Khalil, H. A. A. (1983). Resistance of some plant cultivars to root-knot nematodes, *Meloidogyne* spp. *Nematologia Mediterranea*, 11, 189-192.
- Ibrahim, I. K. A., Mokbel, A. A. & Handoo, Z. A. (2010). Current status of phytoparasitic nematodes and their host plants in Egypt. *Nematropica*, 40, 239-262.

- Ibrahim, I. K. A. & Handoo, Z. A. (2018). Pathogenicity and control of *Meloidogyne incognita* on rice in Egypt. *Pakistan Journal of Nematology*, 36, 123-129.
- Ibrahim, I. K. A., Shahda, W. T. & Dawood, O. A. I. (1999). Pathogenicity and control of *Meloidogyne incognita* on eggplant. *Nematologia Mediterranea*, 27, 31-33.
- Ibrahim, I. K. A., Handoo, Z. A., & El-Sherbiny, A. A. (2000). A survey of phytoparasitic nematodes on cultivated and non-cultivated plants in northwestern Egypt. *Supplement to the Journal of Nematology*, 32 (4S), 478-485.
- Ibrahim, I. K. A., Basyony, A. B. A., Handoo, Z. A. & Chitwood, D. J. (2013). Pathogenicity and control of *Heterodera schachtii* and *Meloidogyne* spp. on some cruciferous plant cultivars. *International Journal of Nematology*, 23, 73-81.
- Ismail, A. E. & Eissa, M. F. M. (1993). Plant-parasitic nematodes associated with ornamental palms in three botanic gardens in Egypt. *Pakistan Journal of Nematology*, 11, 53-59.
- Ivanova, B. P. (1984). Plant parasitic nematodes on flax plantation in Belorussian SSR. *Zashchita Rastenii Minsk*, 9, 37-45.
- Jabeen, S., Bilqees, F. M., Khan, A. & Khatoon, N. (1996). Pathogenicity of *Meloidogyne javanica* on banana in Pakistan. *Proceedings of Parasitology*, 21, 11-16.
- Jonathan, E. I., Barker, K. R. & Abd-El-Aleem, F. F. (1999). Host status of banana for four major species and host races of *Meloidogyne*. *Nematologia Mediterranea*, 27, 123-125.
- Marcelion, Z. C. & Davide, R. G. (1967). Pathogenicity and identity of root-knot nematodes on five varieties of banana. *Philippine Agriculturist*, 2, 241-251.
- Marull, J. & Pinochet, J. (1991). Host suitability of *Prunus* to four *Meloidogyne* species and *Pratylenchus vulnus* in Spain. *Nematropica*, 21, 185-195.
- Marull, J., Pinochet, J., Verdejo, S., & Soler, A. (1991). Reaction of *Prunus* rootstocks to *Meloidogyne incognita* and *M. arenaria* in Spain. *Journal of Nematology*, 23, 564-569.
- Prasad, D. & Khan, E. (1990). Faunistic survey and ecology of plant parasitic nematodes infecting rabi oil-seed crops. *Current Nematology*, 1, 73-76.
- Philis, I. (1989). Resistance of peach seedlings to root-knot nematode attack. *International Nematology Network Newsletter*, 3, 3-5.
- Pinochet, J., Angles, M., Dalmau, E., Fernandez, C. & Felipe, A. (1996). *Prunus* rootstocks evaluation to root-knot and lesion nematodes in Spain. *Journal of Nematology*, 28, 616-623.
- Pinochet, J., Carmen-Jaizme, J. M., Fernandez, C., Joumet, M. & De-Waele, D. (1998). Screening bananas for root-knot (*Meloidogyne* spp.) and lesion nematode (*Pratylenchus goodeyi*) resistance for the Canary Islands. *Fundamental and Applied Nematology*, 21, 17-23.
- Sarah, J. L. (1989). Banana nematodes and their control in Africa. *Nematropica*, 19, 199-216.
- Starr, J. L. & Martyn, R. D. (1991). Reaction of cotton cultivars to *Fusarium* wilt and root-knot nematode. *Nematropica*, 21, 51-58.
- Taylor, A. L. & Sasser, J. N. (1978). *Biology, identification and control of root-knot nematodes (Meloidogyne spp.)*. North Carolina State University Graphics, Raleigh, N.C., U.S.A. 111pp.
- Wendt, K. R. & Greeff, M. S. (1991). Response of selected cotton breeding lines and cultivars to the nematode *Meloidogyne incognita* race 4. *Phytophylactica*, 32, 291-294.
- Youssef, M. M. A. & Eissa, M. F. M. (1994). Population dynamics of certain nematodes associated with date palm in Egypt. *Afro-Asian Journal of Nematology*, 4, 68-72.
- Wrather, J. A., Niblack, T. L. & Milam, M. R. (1992). Survey of plant-parasitic nematodes in Missouri cotton fields. *Journal of Nematology*, 24, 779-7782.
- Zid, A. M. (1996). *Studies on root-knot nematodes attacking sunflower and Flax*. M.Sc. Thesis Faculty of Agriculture, Alexandria University, Alexandria, Egypt. 79 pp.